ORGANIC LETTERS

2002 Vol. 4, No. 21 3561-3564

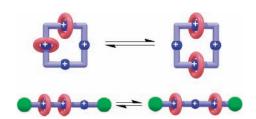
Translational Isomerism in a [3]Catenane and a [3]Rotaxane

Sheng-Hsien Chiu, Arkadij M. Elizarov, Peter T. Glink, and J. Fraser Stoddart*

Department of Chemistry and Biochemistry, University of California, Los Angeles, 405 Hilgard Avenue, Los Angeles, California 90095-1569 stoddart@chem.ucla.edu

Received July 3, 2002

ABSTRACT



Post-assembly covalent modification using Wittig chemistry of [2]rotaxane ylides, wherein NH_2^+ centers in the dumbbell-shaped components are recognized by dibenzo[24]crown-8 (DB24C8) rings, has afforded a [3]catenane and a [3]rotaxane with a precise and synthetically prescribed shortage of DB24C8 rings. The nondegenerate pairs of translational isomers present in both of these interlocked molecular compounds provide the fundamental platform on which to construct sensory devices and nanochemomechanical systems.

Because of their ability to flip between two or more states when external stimuli induce relative movements of their noncovalently interacting components, mechanically interlocked molecules, such as catenanes¹ and rotaxanes,² hold considerable promise³ for the fabrication of actuators, amplifiers, motors, sensors, and switches on the nanoscale level.⁴ For this reason, there is a continual need to develop

(1) Catenanes are molecules comprising two or more interlocked rings. For recent examples, see: (a) Chambron, J.-C.; Sauvage, J.-P.; Mislow, K.; De Cian, A.; Fischer, J. *Chem. Eur. J.* **2001**, 7, 4085–4096. (b) Hori, A.; Kumazawa, K.; Kusukawa, T.; Chand, D. K.; Fujita, M.; Sakamoto, S.; Yamaguchi, K. *Chem. Eur. J.* **2001**, 7, 4142–4149. (c) Park, K.-M.; Kim, S.-Y.; Heo, J.; Whang, D.; Sakamoto, S.; Yamaguchi, K.; Kim, K. *J. Am. Chem. Soc.* **2002**, *124*, 2140–2147.

(2) Rotaxanes are molecules in which one or more rings are trapped on the rod section of a dumbbell-shaped component by bulky stoppers. For recent examples, see: (a) Bryant, W. S.; Guzei, L. A.; Rheingold, A. L.; Gibson, H. W. Org. Lett. 1999, I, 47–50. (b) Seel, C.; Vögtle, F. Chem. Eur. J. 2000, 6, 21–24. (c) Tachibana, Y.; Kihara, N.; Ohga, Y.; Takata, T. Chem. Lett. 2000, 806–807. (d) Mahoney, J. M.; Shukla, R.; Marshall, R. A.; Beatty, A. M.; Zajicek, J.; Smith, B. D. J. Org. Chem. 2002, 67, 1436–1440. (e) Asakawa, M.; Brancato, G.; Fanti, M.; Leigh, D. A.; Shimizu, T.; Slawin, A. M. Z.; Wong, J. K. Y.; Zerbetto, F.; Zhang, S. J. Am. Chem. Soc. 2002, 124, 2939–2950. (f) Chiu, S.-H.; Stoddart, J. F. J. Am. Chem. Soc. 2002, 124, 4174–4175. (g) Andersson, M.; Linke, M.; Chambron, J.-C.; Davidsson, J.; Heitz, V.; Hammarstroem, L.; Sauvage, J.-P. J. Am. Chem. Soc. 2002, 124, 4347–4362. (h) Stanier, C. A.; Alderman, S. J.; Claridge, T. D. W.; Anderson, H. L. Angew. Chem., Int. Ed. 2002, 41, 1769–1772.

(3) (a) Balzani, V.; Credi, A.; Raymo, F. M.; Stoddart, J. F. *Angew. Chem., Int. Ed.* **2000**, *39*, 3348–3391. (b) Ballardini, R.; Balzani, V.; Credi, A.; Gandolfi, M. T.; Venturi, M. *Acc. Chem. Res.* **2001**, *34*, 445–455.

new and efficient methods to prepare catenanes and rotaxanes with multiple recognition sites and to investigate the translational isomerism involving their interactive components. Previously, we have demonstrated,⁵ by employing the Wittig reaction, that benzylic triphenylphosphonium-stoppered [2]rotaxanes,⁶ in which the NH₂⁺ recognition sites on the dumbbell-shaped components are encircled⁷ by dibenzo-[24]crown-8 (DB24C8), are convenient building blocks for the one-pot synthesis of a [4]molecular necklace⁸ and a branched [4]rotaxane.⁹ Here, we report how a [2]rotaxane,⁶ carrying one such stopper, may be grafted onto and crafted

^{(4) (}a) Collier, C. P.; Wong, E. W.; Belohradsky, M.; Raymo, F. M.; Stoddart, J. F.; Kuekes, P. J.; Williams, R. S.; Heath, J. R. *Science* **1999**, 285, 391–394. (b) Collier, C. P.; Mattersteig, G.; Wong, E. W.; Luo, Y.; Beverly, K.; Sampaio, J.; Raymo, F. M.; Stoddart, J. F.; Heath, J. R. *Science* **2000**, 289, 1172–1175. (c) Luo, Y.; Collier, C. P.; Jeppesen, J. O.; Nielsen, K. A.; Delonno, E.; Ho, G.; Perkins, J.; Tseng, H.-R.; Yamamoto, T.; Stoddart, J. F.; Heath, J. R. *ChemPhysChem* **2002**, *3*, 519–525.

⁽⁵⁾ Rowan, S. J.; Stoddart, J. F. J. Am. Chem. Soc. 2000, 122, 164–165.

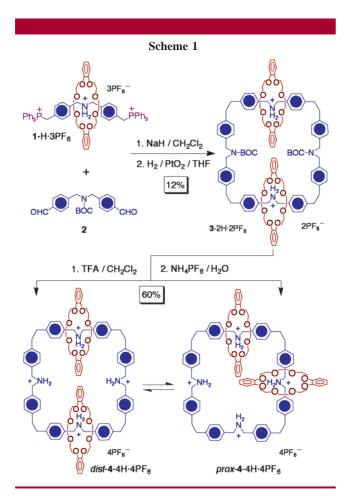
⁽⁶⁾ Rowan, S. J.; Cantrill, S. J.; Stoddart, J. F. Org. Lett. 1999, 1, 129–132.

^{(7) (}a) Glink, P. T.; Schiavo, C.; Stoddart, J. F.; Williams, D. J. *Chem. Commun.* **1996**, 1483–1490. (b) Fyfe, M. C. T.; Stoddart, J. F. *Coord. Chem. Rev.* **1999**, *183*, 139–155. (c) Hubin, T. J.; Kolchinski, A. G.; Vance, A. L.; Busch, D. H. *Adv. Supramol. Chem.* **1999**, *5*, 237–357. (d) Cantrill, S. J.; Pease, A. R.; Stoddart, J. F. *J. Chem. Soc.*, *Dalton Trans.* **2000**, 3715–3734

⁽⁸⁾ Chiu, S.-H.; Rowan, S. J.; Cantrill, S. J.; Ridvan, L.; Ashton, P. R.; Garrell, R. L.; Stoddart, J. F. *Tetrahedron* **2002**, *58*, 807–814.

into (i) a [3]catenane and (ii) a [3]rotaxane with *four* and *three* NH₂+ recognition sites, respectively, for *two* DB24C8 rings. Thus, while the translational isomerism exhibited in solution by the [3]catenane is reminiscent of a model circular railroad with four stations and two trains, the [3]rotaxane resembles a model straight railroad with three stations for two trains. We also demonstrate that in both the [3]catenane and [3]rotaxane the proximal (*prox*) and distal (*dist*) isomers are almost equally populated in solution, a property that could be exploited in the development of sensory devices and nanochemomechanical systems.

Previously, we have reported⁸ an unsuccessful attempt to make a [3]catenane from 1-H·3PF₆ (Scheme 1) by subjecting



it to a bis-Wittig olefination with a benzyloxymethoxy-protected 2,5-dihydroxyterephthalaldehyde derivative. We suspect that our failure to isolate a [3]catenane was the result of steric overcrowding of the two DB24C8 rings, a situation that disfavors macrocyclization and possibly also encourages subsequent hydrogenolysis, rather than hydrogenation, when the crude complex mixture of olefins is treated with hydrogen in the presence of a catalyst. Consequently, we decided to use a dialdehyde with a longer spacer unit separating the two formyl groups and also to employ it to introduce additional NH₂⁺ recognition sites into the [3]catenane.

Dialdehyde¹⁰ 2 not only fulfills these requirements but its Boc-protected amino function acts also as a temporary stopper to prevent the extrusion of the DB24C8 rings during the Wittig olefinations: removal of protecting groups after cyclization unmasks extra recognition sites for the crown ethers. Thus, when an equimolar mixture of 1-H·3PF₆ and 2 was reacted (Scheme 1) in the presence of base (NaH/ CH₂Cl₂) under high dilution conditions, the [3]catenane 3-2H·2PF₆ was isolated in 12% yield, following hydrogenation (H₂/PtO₂/THF) and chromatography (SiO₂: CH₂Cl₂/ MeCN, 9:1). No higher order catenanes were observed¹¹ by mass spectrometry, which revealed peaks at m/z 2137 and 1991 corresponding to [3-2H·PF₆]⁺ and [3-H]⁺, respectively. The molecular train set 4-4H•4PF₆ was obtained in 60% yield after removal (TFA/CH₂Cl₂) of the Boc protecting groups and counterion exchange (NH₄PF₆/H₂O). We investigated the distribution in solution by NMR spectroscopy of the two translational isomers of the molecular train set—one (prox-4-4H·4PF₆) in which the two DB24C8 rings are proximal and the other (dist-4-4H·4PF₆) where they are distal. A previously reported¹² molecular train set—comprising two tetracationic cyclophanes encircling a four-station π -electronrich macrocyclic polyether-was found by ¹H NMR spectroscopy to exist in solution as only the dist translational isomer, presumably in order to minimize electrostatic repulsions between the two tetracationic cyclophanes. In the case of 4-4H·4PF₆, however, there is no such repulsion and so the prox and dist translational isomers are expected to be of similar energies.¹³ On account of the overlapping of signals in the ¹H NMR spectrum, ¹³C NMR spectroscopy was used to investigate the proportions of the two translational isomers. Because of the rapid rotation of the DB24C8 rings of 4-4H. 4PF₆ in the vicinity of their NH₂⁺ recognition sites and the C_{2v} and D_{2h} symmetries of the *prox* and *dist* translational isomers, respectively, we expected to observe one set of signals in the ¹³C NMR spectra of each of these translational isomers. The ¹³C NMR spectrum of 4-4H•4PF₆ in CD₂Cl₂ shows two distinct sets of signals (a, c, d, e in Figure 1) arising from the DB24C8 rings.¹⁴ Although the signals for the tetracationic macrocycle are more complicated than those

3562 Org. Lett., Vol. 4, No. 21, 2002

⁽⁹⁾ Chiu, S.-H.; Rowan, S. J.; Cantrill, S. J.; Stoddart, J. F.; White, A. J. P.; Williams, D. J. *Chem. Eur. J.*, in press.

^{(10) (}a) Ashton, P. R.; Fyfe, M. C. T.; Glink, P. T.; Menzer, S.; Stoddart, J. F.; White, A. J. P.; Williams, D. J. *J. Am. Chem. Soc.* **1997**, *119*, 12514–12524. (b) Chiu, S.-H.; Pease, A. R.; Stoddart, J. F.; White, A. J. P.; Williams, D. J. *Angew. Chem.*, *Int. Ed.* **2002**, *41*, 270–274.

⁽¹¹⁾ The only byproducts in this synthesis were compounds, possibly oligomeric rotaxanes, that could not be moved off the baseline during TLC analysis.

⁽¹²⁾ Ashton, P. R.; Brown, C. L.; Chrystal, E. J. T.; Parry, K. P.; Pietraszkiewicz, M.; Spencer, N.; Stoddart, J. F. *Angew. Chem., Int. Ed. Engl.* **1991**, *30*, 1042–1045.

⁽¹³⁾ The symmetry numbers of the *prox* and *dist* isomers of this compound differ by a factor of 2. Thus, a statistical mixture of the translational isomers should exist as a 2:1 ratio of the *prox* and *dist* isomers. See: Bailey, W. F.; Monahan, A. S. *J. Chem. Ed.* **1978**, *55*, 489–493.

⁽¹⁴⁾ In CD₃CN, the resolution in the ¹³C NMR spectra of these two sets of signals for the DB24C8 rings was poorer when compared with those obtained in CD₂Cl₂. Additionally, increasing the temperature reduced the signal separation and decreasing the temperature increased it. These observations suggest that the resolution of the signals is related to the rate of interconversion between the *prox* and *dist* isomers. At high temperatures or in more-polar solvents, less energy is required to break the hydrogen bonding interactions, both influences which will lower the activation free energy for the shutting process. Thus, the rate of exchange between the two translational isomers is increased and results in poorer resolution of the signals.

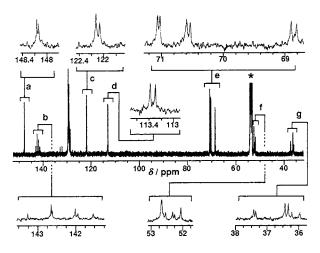
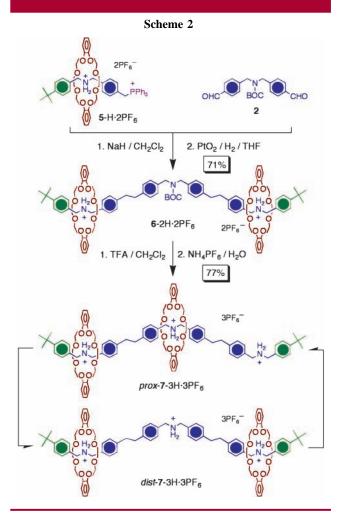


Figure 1. Partial ¹³C NMR spectra (125 MHz, CD₂Cl₂, 298 K) of **4**-4H·4PF₆ displaying a ca. 1:1 mixture of *prox* and *dist* isomers interconverting slowly. Each carbon atom of the DB24C8 subunits (a, c, d, and e) appears as two signals. The signals of the ArCH₂-CH₂Ar units (g) and one of the aromatic carbon atoms (b) of the tetracationic macrocycle reveal the expected six different environments they each encounter (in 1:1:1:1:2:2 ratios), but only five of them are resolved (in a 3:1:1:1:2 ratio) in the case of the CH₂N⁺ units (f).

for the crown ether rings, they are still consistent with the symmetries of the two translational isomers. There are two different environments for the CH₂N⁺ carbon atoms in dist-4-4H·4PF₆ and four different environments for them in prox-**4**-4H•4PF₆. In an approximately 1:1 mixture of the two translational isomers, one would predict that a total of six different environments would result in six peaks for these carbon atoms in approximately a 1:1:1:1:2:2 ratio. This situation manifests itself in the signal (Figure 1f) observed for the CH₂N⁺ carbon atoms, but not all of the six signals are resolved and a 3:1:1:1:2 ratio of them is the outcome. The signals (Figure 1g) for the methylene carbon atoms of the ArCH₂CH₂Ar units and for one (Figure 1b) of the aromatic carbon atoms do, however, display all of the six signals in well-resolved 1:1:2:2:1:1 and 1:2:1:2:1:1 ratios, respectively. Because of the almost equal concentrations of the two translational isomers in solution, ¹³ prox-4-4H•4PF₆ must be disfavored by a weak steric effect. Attempts to observe coalescence of the signals in ¹³C NMR spectra and hence determine the activation barrier for interconversion between the two translational isomers—were thwarted by the fact that the [3]catenane 4-4H·4PF₆ was found¹⁵ to be thermally unstable.

A similar synthetic strategy was employed to make the [3]rotaxane $7\text{-}3H\text{-}3PF_6$ comprising a dumbbell-shaped trication in which three NH_2^+ centers are encircled by two DB24C8 rings. This [3]rotaxane was isolated (Scheme 2) after Wittig olefination of the benzylic monotriphenylphosphonium-stoppered [2]rotaxane $5\text{-}H\text{-}2PF_6$ with the dialdehyde



2, followed by removal of the Boc protecting group on the central amino function in 6-2H·2PF₆ to give the molecular train set 7-3H·3PF₆ as a mixture of translational isomers. Once again, the ¹³C NMR spectrum (125 MHz) at ambient temperature in CD₂Cl₂ of the [3]rotaxane shows two sets of signals of roughly equal intensities for the DB24C8 rings, an observation that is consistent with there being two translational isomers of 7-3H·3PF₆—one (prox-7-3H·3PF₆) in which the two DB24C8 rings are proximal and another (dist-7-3H·3PF₆) in which they are distal. Since the differences in the chemical shifts between the two sets of signals are small, we were unable to extract meaningful data from the ¹³C NMR spectra. Fortunately, however, the ¹H NMR spectra of 7-3H·3PF₆ display distinctly different signals for the protons of the terminal tert-butyl groups in the two translational isomers. In both CD₂Cl₂ and CD₃SOCD₃, the chemical shift of the signal for a tert-butyl substituent on a phenyl group adjacent to an NH₂⁺ center appears further upfield when that NH₂⁺ center is encircled by a DB24C8 ring than when it is not, presumably because of shielding effects by the crown ether's catechol units. In dist-7-3H. 3PF₆ both tert-butyl groups are homotopic and one singlet is expected in the upfield region. In prox-7-3H·3PF₆, however, they are heterotopic and so two singlets of equal intensities are expected—one relatively upfield and the other

Org. Lett., Vol. 4, No. 21, 2002

⁽¹⁵⁾ Although there is no obvious reason for its instability, we observed signals for free DB24C8 in the ^{13}C NMR spectrum of 4-4H·4PF₆ when it was heated above 60 °C in CD₃CN.

relatively downfield-for this translational isomer. Not unexpectedly, perhaps, the more upfield of these signals for both dist- and prox-7-3H-3PF₆ overlap with each other. Thus, the relative concentrations-which are related to the equilibrium constant (K_{eq})—of the *dist/prox* translational isomers of 7-3H·3PF₆ can be determined by taking the ratio of the integral of the more upfield signal (minus the integral of the downfield one) and double the integral of the more-downfield signal. Using this method, the value of K_{eq} was determined to be 0.53 in both CD₂Cl₂ and CD₃SOCD₃ at 298 K. Thus, prox- is more stable 12 than dist-7-3H·3PF₆, in both nonpolar and polar solvents, by ca. 0.4 kcal mol⁻¹ at room temperature. To determine the enthalpic and entropic contributions to this free energy difference between these two translational isomers, we studied the equilibrium at different temperatures. The plot (Figure 2) of the free energy difference (ΔG°) as a

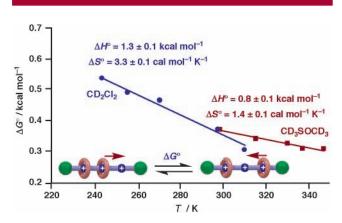


Figure 2. Plot of ΔG° vs T for the equilibrium between prox-7-3H·3PF₆ and dist-7-3H·3PF₆ in CD₂Cl₂ (blue circle) and CD₃-SOCD₃ (red box) determined by 500 MHz ¹H NMR spectroscopy. The slope and intercept of each line of best fit give the values of ΔS° and ΔH° , respectively, from the equation $\Delta G^{\circ} = \Delta H^{\circ} - T\Delta S^{\circ}$.

function of temperature (T) gave straight lines, from which ΔH° and ΔS° were calculated. In both solvents, prox-7-3H•3PF₆ is the more enthalpically stable and dist-7-3H•3PF₆ the more entropically stable. ¹⁶

To obtain a value for the activation energy required to shuttle a DB24C8 ring between $\mathrm{NH_2}^+$ stations in 7-3H·3PF₆, we investigated the coalescence of the signals of the *tert*-butyl groups by variable-temperature ¹H NMR spectroscopy. In CD₃SOCD₃, the signals for the *tert*-butyl groups coalesced at 400 K (Figure 3). Although varying the temperature does affect the equilibrium between the two translational isomers, their ratio remains close to 1:1 over the range 380–420 K. As such, we can employ¹⁷ the approximate rate expression $(k_c = \pi \Delta \nu / \sqrt{2})$ to calculate the rate (k_c) of shuttling at the

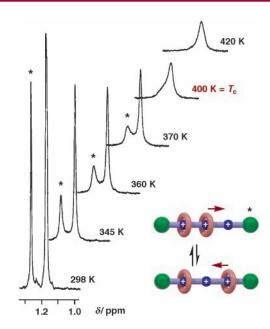


Figure 3. Partial variable-temperature 500 MHz ¹H NMR spectra displaying the coalescence of the *tert*-butyl signals of the [3]rotaxane **7**-3H·3PF₆ in CD₃SOCD₃. The signal marked with the asterisk corresponds to the resonance of the *tert*-butyl protons adjacent to an unoccupied NH₂⁺ center.

coalescence temperature to be ca. 105 s^{-1} , a value of $k_{\rm c}$ that corresponds to an energy barrier ($\Delta G_{\rm c}^{\dagger}$) of ca. 20 kcal mol^{-1} . These values are very similar to those determined ($k_{424\rm K}=98 \text{ s}^{-1}$, $\Delta G_{424\rm K}^{\dagger}=21.3 \text{ kcal mol}^{-1}$) for a simple two-station [2]rotaxane¹⁸ comprised of a DB24C8 ring and a dumbbell-shaped dication in which the NH₂⁺ centers are separated by a p-xylyl unit.

We have established that Wittig chemistry can be employed to advantage to convert appropriately functionalized [2]rotaxanes into a [3]catenane and a [3]rotaxane, both of which can exist as two slowly interconverting translational isomers on the NMR time scale. These nondegenerate interlocked molecules complement the dual-mode co-conformational switching in catenanes¹⁹ incorporating bipyridinium and dialkylammonium sites and avail us of yet another opportunity to develop molecular-level machines.³

Acknowledgment. We thank the Petroleum Research Fund, administered by the American Chemical Society, for generous financial support. This material is based upon work supported by the National Science Foundation under equipment grant no. CHE-9974928.

Supporting Information Available: Synthesis and characterization of all the new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

OL026464E

3564 Org. Lett., Vol. 4, No. 21, 2002

⁽¹⁶⁾ Statistically (ref 13), the dist/prox ratio is expected to be 0.5, suggesting—from consideration of the "trivial" entropy of the system—that the prox isomer should be the more entropically stable (by ca. 1.4 cal mol⁻¹ K⁻¹). The observation that the dist isomer is the more entropically stable suggests that symmetry factors alone do not explain the observed dist/prox ratio of 0.53.

⁽¹⁷⁾ Sutherland, I. O. Annu. Rep. NMR Spectrosc. 1971, 4, 71-235.

⁽¹⁸⁾ Cao, J.; Fyfe, M. C. T.; Stoddart, J. F.; Cousins, G. R. L.; Glink, P. T. J. Org. Chem. **2000**, *65*, 1937–1946.

⁽¹⁹⁾ Ashton, P. R.; Baldoni, V.; Balzani, V.; Credi, A.; Hoffmann, H. D. A.; Martínez-Díaz, M.-V.; Raymo, F. M. R.; Stoddart, J. F.; Venturi, M. *Chem. Eur. J.* **2001**, *7*, 3482–3493.